Streptomonospora halophila sp. nov., a halophilic actinomycete isolated from a hypersaline soil

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An actinomycete strain, designated YIM 91355<sup>T</sup>, was isolated from a soil sample collected from a hypersaline soil in Xinjiang Province, China, and its taxonomic position was investigated by using a polyphasic approach. The strain grew well on most media tested and no diffusible pigment was produced. The aerial mycelium of this organism was well developed but not fragmented and, at maturity, formed short chains of spores. Substrate mycelium was branched with nonfragmenting hyphae and single, oval to round spores were formed, borne on sporophores. Strain YIM 91355<sup>T</sup> contained *meso*-diaminopimelic acid as the diagnostic amino acid. The purified cell-wall hydrolysate contained galactose. The predominant menaquinones were MK-10(H<sub>8</sub>) (43.2%), MK-10(H<sub>6</sub>) (17.7%) and MK-11(H<sub>8</sub>) (10.4%). Phospholipids were phosphatidylglycerol, diphosphatidylglycerol, phosphatidylinositol mannosides, phosphatidylcholine, phosphatidylinositol and an unidentified phospholipid. The major fatty acids were i-C<sub>16:0</sub>, ai-C<sub>17:0</sub>, 10-methyl C<sub>17:0</sub> and 10-methyl C<sub>18:0</sub>. The G+C content of the genomic DNA was 72.1 mol%. Phylogenetic analysis indicated that the isolate belongs to the genus Streptomonospora of the family Nocardiopsaceae. Based on phenotypic and genotypic data, it is concluded that strain YIM 91355<sup>T</sup> represents a novel species of the genus *Streptomonospora*, for which the name Streptomonospora halophila sp. nov. (type strain YIM 91355<sup>T</sup> =DSM 45075<sup>T</sup> =KCTC 19236<sup>T</sup>) is proposed.

The genus Streptomonospora was first established by Cui et al. (2001) with the type species Streptomonospora salina and it represents a group of strictly halophilic filamentous actinomycetes that form a distinct branch in the 16S rRNA gene phylogenetic tree adjacent to the genera Nocardiopsis and Thermobifida of the family Nocardiopsaceae (Stackebrandt et al., 1997). Subsequently, Li et al. (2003) described another species, Streptomonospora alba, and emended the description of the genus Streptomonospora. During a biodiversity and taxonomic study on halophilic filamentous actinomycetes, numerous Streptomonosporalike or Nocardiopsis-like strains have been isolated from the same hypersaline habitat as S. salina YIM  $90002^{T}$  and S. alba YIM 90003<sup>T</sup>, i.e. close to Aiding Lake in Xinjiang Province, north-west China. For rapid identification of Streptomonospora species, genus-specific PCR amplification

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was introduced according to the method described by Zhi *et al.* (2006). As a result, a third halophilic *Streptomonospora* isolate, strain YIM  $91355^{T}$ , attracted our attention and was investigated using a polyphasic approach to determine its taxonomic position.

Strain YIM 91355<sup>T</sup> was isolated and maintained on modified ISP 5 medium (0.1 %  $K_2$ HPO<sub>4</sub>, 0.1 % L-asparagine, 1.0 % glycerol, 0.5 % yeast extract, 0.5 % KNO<sub>3</sub>, 10.0 % NaCl, 2.0 % agar, pH 7.0). Other media used for examining cultural characteristics were inorganic salts/starch agar, Czapek's agar, potato agar, nutrient agar, yeast extract/malt extract agar and oatmeal agar (Shirling & Gottlieb, 1966), all of which were supplemented with 10 % (w/v) NaCl; see data in Table 1. Strain YIM 91355<sup>T</sup> grew well on most media, especially on modified ISP 5 medium. No diffusible pigments were produced on any of the media tested.

Morphological characteristics of the strain were observed by light microscopy (model BH 2; Olympus) and scanning electron microscopy (JSM5600LV; JEOL) after 7 days

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Abbreviation: DAP, diaminopimelic acid.

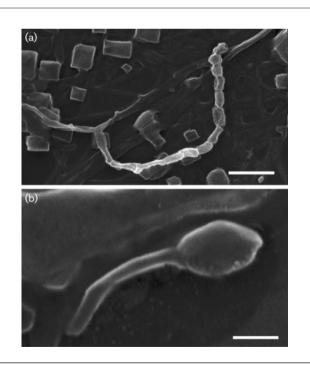
The GenBank/EMBL/DDBJ accession number for the 16S rRNA gene sequence of strain YIM  $91355^{T}$  is EF423989.

#### **Table 1.** Cultural characteristics of strain YIM 91355<sup>T</sup>

Diffusible pigments were not produced on any of the media listed. All media contained 10 % (w/v) NaCl,	
pH 7.0. Colours were taken from the ISCC-NBS colour charts (standard samples, no. 2106) (Kelly, 1964).	

Medium	Growth	Colour of colonies	
		Aerial mycelium	Substrate mycelium
Czapek's agar	Moderate	Yellow-white	Pale yellow
Nutrient agar	Poor	Yellow-white	Pale yellow
Potato agar	Good	Yellow-white	Pale yellow
Modified ISP 5 medium	Good	Yellow-white	Pale yellow
Inorganic salts/starch agar	Moderate	White	Pale yellow-white
Yeast extract/malt extract agar	Good	Yellow-white	Yellowish brown
Oatmeal agar	Good	Yellow-white	Pale yellow

growth on modified ISP 5 containing 10% NaCl (w/v) at 37 °C. Investigations of 7-day-old cultures of strain YIM 91355<sup>T</sup> revealed that the isolate shared the same morphological characteristics as members of the genus *Streptomonospora*. The aerial mycelium was well developed but not fragmented and, at maturity, it formed spore chains that were straight to flexuous. Spores were oval to cylindrical (0.4–0.5 × 0.8–1.0 µm) and non-motile (Fig. 1a). Substrate mycelium was extensively branched with nonfragmenting hyphae. Single spores, oval to round and 0.8– 1.0 µm in diameter, were borne on sporophores of substrate mycelium (Fig. 1b). The colours of substrate



**Fig. 1.** Scanning electron micrographs of strain YIM  $91355^{T}$  grown on modified ISP medium 5 for 7 days at 37 °C showing a spore chain on aerial mycelium (a; bar, 2 µm) and a single spore on substrate mycelium (b; bar, 500 nm).

and aerial mycelia were determined by comparison with chips from ISCC-NBS colour charts (Kelly, 1964).

Growth was assessed at 4, 10, 20, 28, 37, 45, 55, 60 and 65 °C; optimum growth was observed at 37 °C. pH and NaCl tolerance was examined as described by Tang *et al.* (2003). Other physiological features and carbon source utilization tests were observed on media commonly used for the characterization of *Streptomyces* species (Shirling & Gottlieb, 1966; Williams *et al.*, 1989) supplemented with 10 % (w/v) NaCl. Detailed physiological and biochemical characteristics of the isolate are given in Table 2 and in the species description.

The cell wall amino acids were analysed by using TLC as described by Lechevalier & Lechevalier (1980). Cell walls contained meso-diaminopimelic acid (meso-DAP). Sugar analysis of cell walls followed procedures described by Staneck & Roberts (1974); the hydrolysate of purified cell walls of strain YIM 91355<sup>T</sup> contained galactose. Polar lipids were extracted, examined by two-dimensional TLC and identified using previously described procedures (Minnikin et al., 1979). Strain YIM 91355<sup>T</sup> possessed diphosphatiphosphatidylglycerol, phosphatidylinositol dylglycerol, mannosides, phosphatidylcholine, phosphatidylinositol and an unidentified phospholipid. The strain was cultivated in modified ISP 5 medium for menaquinone analysis. Menaquinones were extracted as described by Collins et al. (1977) and were analysed by HPLC (Kroppenstedt, 1982). The predominant menaquinones were MK-10( $H_8$ ) (43.2%), MK-10(H<sub>6</sub>) (17.7%) and MK-11(H<sub>8</sub>) (10.4%); MK-9(H<sub>4</sub>) (1.1%), MK-9(H<sub>6</sub>) (1.7%), MK-9(H<sub>8</sub>) (3.7%), MK-10 (0.9%), MK-10(H<sub>2</sub>) (2.4%), MK-10(H<sub>4</sub>) (6.8%), MK-11(H<sub>4</sub>) (1.0%), MK-12 (5.2%), MK-11(H<sub>6</sub>) (4.0%) and MK-11(H<sub>10</sub>) (1.8%) were minor components. Cellular fatty acid analysis was performed as described by Sasser (1990) using the Microbial Identification system (MIDI). The fatty acid profile of strain YIM 91355<sup>T</sup> was mainly composed of i-C<sub>16:0</sub> (39.8 %), ai-C<sub>17:0</sub> (12.5 %), 10-methyl  $C_{17:0}$  (10.2 %) and 10-methyl  $C_{18:0}$  (tuberculostearic acid; 11.1%); for comparison, the fatty acid compositions of the type strains of S. salina and S. alba are shown in Table 2.

### **Table 2.** Differential phenotypic characteristics of strain YIM 91355<sup>T</sup> and the type strains of *Streptomonospora* species

Data are from this study, Cui *et al.* (2001) and Li *et al.* (2003). For all three strains, the optimal growth temperature is 37 °C. They have *meso*-DAP as the diagnostic amino acid in the cell wall. All are negative for gelatin liquefaction, milk coagulation and utilization of arabinose, mannitol, raffinose and sucrose. +, Positive; -, negative; NT, not tested.

Characteristic	S. halophila YIM 91355 <sup>T</sup>	S. salina YIM $90002^{T}$	S. alba YIM $90003^{T}$
NaCl concentration for growth (9	%)		
Range	5–20	5–20	5–25
Optimum	10	15	10-15
Milk peptonization	+	_	_
Nitrate reduction	_	-	+
Production of:			
$H_2S$	_	_	NT
Oxidase	_	+	—
Catalase	+	NT	+
Melanin	_	+	—
Amylase	_	+	_
Cellulase	+	_	_
Carbon-source utilization			
Glucose	_	-	+
Inositol	+	-	_
Maltose	+	-	—
Rhamnose	+	-	_
Cell-wall sugar(s)	Galactose	Galactose	Galactose, arabinose
Predominant menaquinones	MK-10(H <sub>8</sub> ) (43.2%), MK-10(H <sub>6</sub> )	MK-10(H <sub>8</sub> ) (39.4%), MK-10(H <sub>6</sub> )	MK-10(H <sub>8</sub> ) (50.6%),
(>10%)*	(17.7%), MK-11(H <sub>8</sub> ) (10.4%)	(22.6 %), MK-10(H <sub>4</sub> ) (11.8 %)	MK-10(H <sub>6</sub> ) (15.8%)
Major phospholipids†	DPG, PG, PC, PIM, PI, PL	PG, PI, PC, 2MPE, PL	PG, PE, PI, DPG, MPE, PS, PC, PL
Major fatty acids‡	i-C <sub>16:0</sub> (39.8%), ai-C <sub>17:0</sub>	$i-C_{15:0}$ (30.1%), $i-C_{16:0}$ (16.5%),	ai-C <sub>17:0</sub> (26.0%), i-C <sub>16:0</sub>
	(12.5 %), 10-methyl $C_{17:0}$ (10.2 %), 10-methyl $C_{18:0}$ (11.1 %)	9-methyl C <sub>16:0</sub> (11.7%), ai-C <sub>17:0</sub> (6.7%)	(25.1 %), $C_{16:0}$ (8.6 %), ai- $C_{15:0}$ (8.0 %)
DNA G+C content (mol%)§	72.1	72.9	74.4

\*Grown on modified ISP 5 medium.

†DPG, Diphosphatidylglycerol; MPE, methylphosphatidylethanolamine; PC, phosphatidylcholine; PE, phosphatidylethanolamine; PG, phosphatidylglycerol; PI, phosphatidylinositol; PIM, PI mannosides; PL, unidentified phospholipid; PS, phosphatidylserine. ‡Grown on modified TSA.

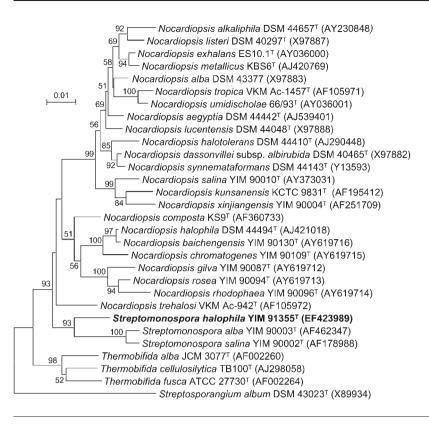
\$Determined using the HPLC method.

The DNA G + C content of strain YIM 91355<sup>T</sup>, determined by using the HPLC method of Mesbah *et al.* (1989), was 72.1 mol%.

Extraction and amplification of genomic DNA for 16S rRNA gene sequence analysis were carried out as described by Li *et al.* (2007). The nearly full-length 16S rRNA gene sequence of strain YIM 91355<sup>T</sup> was determined by direct sequencing of the PCR-amplified 16S rRNA gene. Multiple alignments with sequences of related taxa in the family *Nocardiopsaceae* were implemented by using CLUSTAL\_X (Thompson *et al.*, 1997). 16S rRNA gene similarity values were calculated by using a web-based tool as described by Chun *et al.* (2007) (http://www.eztaxon.org). A phylogenetic tree (Fig. 2) was reconstructed using the neighbourjoining method of Saitou & Nei (1987) from *K*<sub>nuc</sub> values (Kimura, 1980). Topology of the phylogenetic tree was

evaluated by the bootstrap resampling method of Felsenstein (1985) with 1000 replicates.

In the phylogenetic dendrogram, strain YIM 91355<sup>T</sup> formed a stable clade (supported by 93 % bootstrap value; see Fig. 2) together with the two species of the genus *Streptomonospora*, family *Nocardiopsaceae*, with validly published names, even though the highest 16S rRNA gene similarity was found between strain YIM 91355<sup>T</sup> and *Nocardiopsis halophila* KCTC 9825<sup>T</sup> (96.94 %, 1421 bp). The closest phylogenetic relationship was observed between strain YIM 91355<sup>T</sup> and members of the genus *Streptomonospora*; this observation was supported by morphological and chemotaxonomic data. For example, the isolate shared the morphological characteristics described for members of the genus *Streptomonospora*, the aerial mycelium was well developed but not fragmented, the substrate mycelium was extensively



**Fig. 2.** Phylogenetic dendrogram based on 16S rRNA gene sequence analysis reconstructed from evolutionary distances by using the neighbour-joining method, showing the phylogenetic position of strain YIM 91355<sup>T</sup> within the family *Nocardiopsaceae*. Bootstrap values (only above 50%) of 1000 replicates are shown as percentages. *Streptosporangium album* DSM 43023<sup>T</sup> was used as the outgroup. Bar, 1 substitution per 100 nucleotide positions.

branched with non-fragmenting hyphae and single spores were borne on sporophores of substrate mycelium. In addition, the strain contained galactose as the diagnostic sugar, which is consistent with chemotaxonomic characteristics of members of the genus Streptomonospora, but not members of the genus Nocardiopsis. The 16S rRNA gene sequence similarities between YIM 91355<sup>T</sup> and the type strains of S. salina and S. alba were 96.51 and 96.81%, respectively. Group-specific signature analysis showed that strain YIM 91355<sup>T</sup> and both Streptomonospora species shared identical signatures at positions 229 (G), 508 (C), 811 (U), 461-471 (Y-G), 611-629 (U-U) and 1028-1033 (C–G). Thus, all the above data support the inclusion of the isolate in the genus Streptomonospora (Cui et al., 2001; Li et al., 2003). However, strain YIM 91355<sup>T</sup> differs from the other two species of the genus Streptomonospora in many physiological, chemotaxonomic and biochemical properties; details are shown in Table 2.

Furthermore, it is generally recognized that organisms that display sequence divergence values of >3 % do not belong to the same species (Stackebrandt & Goebel, 1994). Thus, based on the above phenotypic and genotypic results, strain YIM 91355<sup>T</sup> represents a novel species of the genus *Streptomonospora*, for which the name *Streptomonospora* halophila sp. nov. is proposed.

### Description of Streptomonospora halophila sp. nov.

Streptomonospora halophila (ha.lo.phi'la. Gr. n. hals, halos salt; Gr. adj. philos loving; N.L. fem. adj. halophila

salt-loving, referring to the ability to grow at high NaCl concentrations).

Gram-positive and aerobic. Grows well on most test media, but no diffusible pigment is produced. Aerial mycelium and substrate mycelium are well developed but not fragmented on most media. The yellow-white aerial mycelium (white only on inorganic salts/starch agar) forms straight to flexuous spore chains at maturity; spores are oval to cylindrical  $(0.4-0.5 \times 0.8-1.0 \ \mu\text{m})$  and nonmotile. Single, oval to round spores  $(0.8-1.0 \ \mu m \ diameter)$ are borne on sporophores of substrate mycelium. Colours of the substrate mycelium are pale yellow on Czapek's agar, nutrient agar, potato agar, modified ISP 5 medium and oatmeal agar, pale yellow-white on inorganic salts/starch agar and yellowish brown on yeast extract/malt extract medium. The optimum NaCl concentration, growth pH and temperature are 10 % (w/v), pH 7.0 and 37 °C. Ranges of NaCl concentration, pH and temperature for growth are 5-20 %, pH 6.0-9.0 and 20-45 °C. Inositol, maltose and rhamnose can be utilized as sole carbon sources. Positive for milk peptonization, catalase and cellulose, but negative for oxidase, melanin production, amylase, nitrate reduction and production of H<sub>2</sub>S. The diagnostic amino acid is meso-DAP; galactose is the main cell wall sugar. The predominant menaquinones are MK-10(H<sub>8</sub>), MK-10(H<sub>6</sub>) and MK-11(H<sub>8</sub>). Phospholipids are diphosphatidylglycerol, phosphatidylglycerol, phosphatidylcholine, phosphatidylinositol mannosides, phosphatidylinositol and an unidentified phospholipid.

The type strain is YIM  $91355^{T}$  (=DSM  $45075^{T}$  =KCTC  $19236^{T}$ ), isolated from a hypersaline soil in Xinjiang Province, north-west China. The G+C content of DNA of the type strain is 72.1 mol%.

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## References

Chun, J., Lee, J.-H., Jung, Y., Kim, M., Kim, S., Kim, B. K. & Lim, Y. W. (2007). EzTaxon: a web-based tool for the identification of prokaryotes based on 16S ribosomal RNA gene sequences. *Int J Syst Evol Microbiol* 57, 2259–2261.

**Collins, M. D., Pirouz, T., Goodfellow, M. & Minnikin, D. E. (1977).** Distribution of menaquinones in actinomycetes and corynebacteria. *J Gen Microbiol* **100**, 221–230.

Cui, X. L., Mao, P. H., Zeng, M., Li, W. J., Zhang, L. P., Xu, L. H. & Jiang, C. L. (2001). *Streptimonospora salina* gen. nov., sp. nov., a new member of the family *Nocardiopsaceae*. *Int J Syst Evol Microbiol* 51, 357–363.

**Felsenstein, J. (1985).** Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* **39**, 783–791.

Kelly, K. L. (1964). Inter-Society Color Council – National Bureau of Standards Color-Name Charts Illustrated with Centroid Colors. Washington, DC: US Government Printing Office.

**Kimura, M. (1980).** A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J Mol Evol* **16**, 111–120.

**Kroppenstedt, R. M. (1982).** Separation of bacterial menaquinones by HPLC using reverse phase (RP18) and a silver loaded ion exchanger as stationary phases. *J Liq Chromatogr* **5**, 2359–2367.

Lechevalier, M. P. & Lechevalier, H. A. (1980). The chemotaxonomy of actinomycetes. In *Actinomycete Taxonomy*, pp. 227–291. Edited by A. Dietz & D. W. Thayer. Arlington, VA: Society for Industrial Microbiology.

Li, W.-J., Xu, P., Zhang, L.-P., Tang, S.-K., Cui, X.-L., Mao, P.-H., Xu, L.-H., Schumann, P., Stackebrandt, E. & Jiang, C.-L. (2003). *Streptomonospora alba* sp. nov., a novel halophilic actinomycete,

and emended description of the genus *Streptomonospora* Cui *et al.* 2001. *Int J Syst Evol Microbiol* **53**, 1421–1425.

Li, W. J., Xu, P., Schumann, P., Zhang, Y. O., Pukall, R., Xu, L. H., Stackebrandt, E. & Jiang, C. L. (2007). *Georgenia ruanii* sp. nov., a novel actinobacterium isolated from forest soil in Yunnan (China), and emended description of the genus *Georgenia*. Int J Syst Evol Microbiol 57, 1424–1428.

**Mesbah, M., Premachandran, U. & Whitman, W. B. (1989).** Precise measurement of the G+C content of deoxyribonucleic acid by high-performance liquid chromatography. *Int J Syst Bacteriol* **39**, 159–167.

Minnikin, D. E., Collins, M. D. & Goodfellow, M. (1979). Fatty acid and polar lipid composition in the classification of *Cellulomonas*, *Oerskovia* and related taxa. *J Appl Bacteriol* 47, 87–95.

Saitou, N. & Nei, M. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4, 406–425.

Sasser, M. (1990). Identification of bacteria by gas chromatography of cellular fatty acids. USFCC Newsl 20, 16.

Shirling, E. B. & Gottlieb, D. (1966). Methods for characterization of *Streptomyces* species. *Int J Syst Bacteriol* 16, 313–340.

**Stackebrandt, E. & Goebel, B. M. (1994).** Taxonomic note: a place for DNA-DNA reassociation and 16S rRNA sequence analysis in the present species definition in bacteriology. *Int J Syst Bacteriol* **44**, 846–849.

**Stackebrandt, E., Rainey, F. A. & Ward-Rainey, N. L. (1997).** Proposal for a new hierarchic classification system, *Actinobacteria* classis nov. *Int J Syst Bacteriol* **47**, 479–491.

**Staneck, J. L. & Roberts, G. D. (1974).** Simplified approach to identification of aerobic actinomycetes by thin-layer chromatography. *Appl Microbiol* **28**, 226–231.

Tang, S. K., Li, W. J., Wang, D., Zhang, Y. G., Xu, L. H. & Jiang, C. L. (2003). Studies of the biological characteristics of some halophilic and halotolerant actinomycetes isolated from saline and alkaline soils. *Actinomycetologica* 17, 6–10.

Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D. G. (1997). The CLUSTAL\_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res* **25**, 4876–4882.

Williams, S. T., Goodfellow, M. & Alderson, G. (1989). Genus *Streptomyces* Waksman and Henrici 1943, 339<sup>AL</sup>. In *Bergey's Manual of Systematic Bacteriology*, vol. 4, pp. 2452–2492. Edited by S. T. Williams, M. E. Sharpe & J. G. Holt. Baltimore: Williams & Wilkins.

Zhi, X. Y., Tang, S. K., Li, W. J., Xu, L. H. & Jiang, C. L. (2006). New genus-specific primers for the PCR identification of novel isolates of the genus *Streptomonospora*. *FEMS Microbiol Lett* **263**, 48–53.