

***Methylobacterium soli* sp. nov. a methanol-utilizing bacterium isolated from the forest soil**

**Yan-Ru Cao · Qian Wang · Rong-Xian Jin ·
Shu-Kun Tang · Yi Jiang · Wen-Xiang He ·
Hang-Xian Lai · Li-Hua Xu · Cheng-Lin Jiang**

Received: 4 July 2010/Accepted: 20 November 2010/Published online: 9 January 2011
© Springer Science+Business Media B.V. 2011

Abstract A Gram-negative, pink-pigmented, non-spore-forming rod shaped, methanol-utilizing bacterium, strain YIM 48816^T, was isolated from forest soil collected from Sichuan province, China. Strain YIM 48816^T can grow at 4–37°C, pH 5.0–7.0 and 0% NaCl (w/v). Based on 16S rRNA gene sequence similarity studies, it belonged to the genus *Methylobacterium*, and formed a phyletic line. The 16S rRNA gene sequence similarities were 96.2% to *Methylobacterium mesophilicum* DSM 1708^T and 96.0% to *Methylobacterium brachiatum* DSM 19569^T, and the phylogenetic similarities to all other *Methylobacterium* species with validly published names were less than 96.0%. The major menaquinones detected were

Q-10 (97.14%) and Q-9 (2.86%). The major fatty acids were C18:1 ω7c (80.84%). The DNA G + C content was 66.2 mol%. It is apparent from the genotypic and phenotypic data that strain YIM 48816^T belongs to a novel species of the genus *Methylobacterium*, for which the name *Methylobacterium soli* sp. nov. is proposed. The type strain is YIM 48816^T (CCTCC AA 208027^T = KCTC 22810^T).

Keywords *Methylobacterium soli* sp. nov · Soil · 16S rRNA gene

Introduction

The genus *Methylobacterium* was established by Patt et al. (1976). The species of this genus exist in many kinds of habitation such as solid, water and air. Growing on one-carbon and multicarbon compounds is the characteristic of this genus. So far, the genus *Methylobacterium* comprised 34 validly described species (<http://www.bacterio.cict.fr/m/methylobacterium.html>).

Materials and methods

Isolation and maintenance of organism

During the course of studying the relationship between bacterial diversity and the various altitudes

The GenBank accession number for the 16S rRNA gene sequence of strain YIM 48816^T is EU860984.

Y.-R. Cao · W.-X. He · H.-X. Lai
College of Resources and Environment, Northwest A&F University, Yangling, 712100 Xianyang, Shaanxi, China

Y.-R. Cao · Q. Wang · R.-X. Jin · S.-K. Tang ·
Y. Jiang (✉) · L.-H. Xu · C.-L. Jiang
The Key Laboratory for Microbial Resources
of the Ministry of Education, Yunnan University,
650091 Kunming, China
e-mail: jiangyikm@hotmail.com

Y.-R. Cao · Q. Wang · R.-X. Jin · S.-K. Tang ·
Y. Jiang · L.-H. Xu · C.-L. Jiang
Laboratory for Conservation and Utilization of
Bio-Resources, Yunnan Institute of Microbiology,
Yunnan University, 650091 Kunming, China

of the mountain, we got the strain YIM 48816^T from the soil which altitude was 3300 m, collecting from Sichuan, Southwest of China. 2 g of the soil sample (dried for 7 days at room temperature and dry heated for 1 h at 80°C) was suspended in 20 ml sterilized distilled water, then shaken at 180 rpm for 1 h. The solution was spreaded onto ISP media 5 (Shirling and Gottlieb 1966). On the basis of colony morphology, one pink-pigmented strain (YIM 48816^T) were purified and maintained on the medium ISP 2 (Shirling and Gottlieb 1966). The strain was highly odoriferous.

Molecular analysis

Biomass for molecular systematic studies was derived from a medium ISP 2 shake culture incubated at 28°C for 10 days, harvested by centrifugation and washed twice using distilled water. Extraction of chromosomal DNA and PCR amplification of the 16S rRNA gene from the species YIM 48816^T were carried out by established procedures (Li et al. 2007). The 16S rRNA gene sequences of related taxa were obtained from the EzTaxon server (<http://www.eztaxon.org/>) (Chun et al. 2007). Multiple alignments of sequence similarity were carried out using CLUSTAL_X (Thompson et al. 1997). Phylogenetic analyses were inferred using three tree-making algorithms, the neighbour-joining (Saitou and Nei 1987), maximum-likelihood (Felsenstein 1981) and maximum-parsimony (Fitch 1971) methods. The tree topology was performed by the neighbour-joining method using MEGA (Molecular Evolutionary Genetics Analysis) version 3.1 (Kumar et al. 2004), and Kimura two-parameter model (Kimura 1980) was used to calculate the distances. Bootstrap analysis was used to evaluate the tree topology of the neighbour-joining data based on 1000 resamplings (Felsenstein 1985).

Because strain YIM 48816^T showed low levels of 16S rRNA gene sequence similarities with all validly published species of the genus *Methylobacterium*, it was subjected to a polyphasic taxonomic investigation.

Morphological, physiological and biochemical characteristics

The Gram reaction was performed by using the standard Gram reaction and confirmed by using the KOH lysis test method (Cerny 1978). Morphological

characteristics were observed by light microscopy (BH2 microscope; Olympus) after 6 days growth on ISP 2 agar medium at 28°C. Salt tolerance was tested in ISP 2 medium supplemented with 1–10% (w/v) NaCl. The effects of temperature on growth were examined at 0, 4, 10, 15, 20, 28, 37, 45 and 55°C for 14 days. Growth at different pH concentrations (from 4.0 to 10.0 at intervals of 1 pH unit) was assessed after incubation for 2 weeks as described by Xu et al. (2005). Carbon and nitrogen source utilization was tested as described by Gordon et al. (1974). Catalase activity was determined by production of bubbles after the addition of a drop of 3% H₂O₂. Oxidase activity was observed by oxidation of tetramethyl-p-phenylenediamine. Urease activity, H₂S production, nitrate reduction, milk coagulation and peptonization, hydrolysis of gelatin, cellulose, starch and Tweens 20, 40, 60 and 80 were tested as described by Cowan and Steel (1965). Other enzyme activities were determined by using the API ZYM systems (bioMe'rieux) according to the manufacturer's instructions.

Chemotaxonomy

To measure the G + C content of the DNA, it was enzymically degraded into nucleosides and investigated as described by Mesbah et al. (1989), using reversed-phase HPLC. Chemosystematic studies were carried out to establish whether isolate YIM 48816^T had a chemical profile consistent with its assignment to the genus *Methylobacterium*. Cell biomass for quinone analysis was obtained from cultivation in ISP 2 medium shake culture incubated at 28°C for 10 days. The isoprenoid quinone was investigated as described by Komagata and Suzuki (1987), using reversed-phase HPLC. The cellular fatty acids were saponified, methylated and extracted according to the protocol of the Sherlock Microbial Identification System (MIDI), and identified using the Microbial Identification software package (Sasser 1990).

Results and discussion

The 16S rRNA gene sequence of the strain YIM 48816^T was a continuous stretch of 1405 bp. Sequence similarity calculations and phylogenetic

analysis revealed that strain YIM 48816^T was closely related to *M. mesophilicum* DSM 1708^T (Austin and Goodfellow 1979; Green and Bousfield 1983) and *M. brachiatum* DSM 19569^T (Kato et al. 2008), with sequence similarities of 96.2 and 96.0%, respectively. The similarities with respect to all other members of this genus were below 96.0%. A phylogenetic tree was constructed with sequences of representative strains of the genus *Methylobacterium*, and the type strain of *Balneimonas flocculans* TFB^T was used as the outgroup. The phylogenetic tree (Fig. 1) based on the neighbour-joining algorithm showed that strain YIM 48816^T falls within the genus *Methylobacterium*, forming a distinct phyletic line with a high bootstrap resampling value of 72%, which confirmed that the new isolate belonged to *Methylobacterium*, but differentiated from all the recognized species of the genus *Methylobacterium*.

The isolate YIM 48816^T was determined as comprising Gram-negative, and formed pink- to light red-pigmented colonies. Cells of the strain were rods and no spores were observed. Cells of the strain often contained poly-β-hydroxybutyrate or volutin granules (Fig. 2). Strain YIM 48816^T did not grow in the presence of 1% NaCl, it was able to grow at 4–37°C and at pH 5.0–7.0. The optimum

growth occurred at 28°C and pH 7.0 on ISP 2 agar medium. Cells were oxidase- and catalase-positive. The other physiological properties determined for strain YIM 48816^T are given in Table 1 and in the species description.

The genomic DNA G + C content of strain YIM 48816^T was 66.2 mol%. The predominant ubiquinone was Q-10. The primary fatty acid was C_{18:1} ω7c (88.46%). All of these chemical properties are consistent with the classification of the isolate YIM 48816^T in the genus *Methylobacterium*.

Taxonomic conclusion

The 16S rRNA gene sequence, phylogenetic data showed that the isolate YIM 48816^T was a member of the genus *Methylobacterium*. Also, the following characteristics (cell shape, the predominant ubiquinone and some other physiological properties) of strain YIM 48816^T are consistent with its assignment to the genus *Methylobacterium*. Whereas, the low levels of 16S rRNA gene sequence similarity distinguish the novel isolate from its closest neighbours in the genus *Methylobacterium*. Table 1 provides a comparison of the characteristics of strain YIM 48816^T and closely related species of the genus

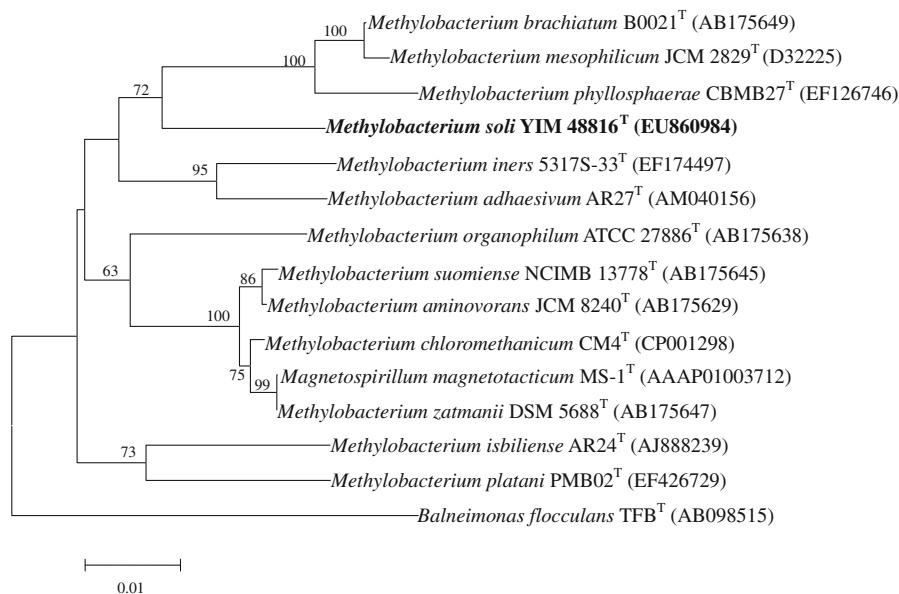


Fig. 1 Neighbour-joining phylogenetic tree based on 16S rRNA gene sequences, showing the position of isolate YIM 48816^T within the *Methylobacterium* genus. Numbers at nodes are bootstrap percentages (based on 1000 resampled datasets);

only values above 50% are given. The sequence of *Balneimonas flocculans* TFB^T (GenBank accession no. AB098515) was used as an outgroup. Bar 0.01 substitutions per nucleotide position



Fig. 2 Transmission electron micrograph showing the general cell morphology of strain YIM 48816^T and the presence of the poly- β -hydroxybutyrate or volutin granules after growth on ISP 2 agar for 6 days at 28°C. Bar 1 μ m

Methylobacterium. The strain YIM 48816^T can use D-fructose, *N,N*-dimethylformamide and L-aspartate as carbon sources, but not L-arabinose or D-xylose;

the closest strains *M. mesophilicum* DSM 1708^T and *M. brachiatum* DSM 19569^T yielded opposite results. Additionally, the fatty acids of *M. mesophilicum* and *M. brachiatum* have traces of C18 :1 ω 9c, which is absent in the strain YIM 48816^T. DNA G + C content of strain YIM 48816^T was lower 3% than the closest strains *M. mesophilicum* DSM 1708^T and *M. brachiatum* DSM 19569^T. These physiological and biochemical characteristics (Table 1) evidently support the strain YIM 48816^T represents a novel species of the genus *Methylobacterium*, for which the name *Methylobacterium soli* sp. nov. is proposed.

Description of *Methylobacterium soli* sp. nov

Methylobacterium soli (so'li. L. neut. gen. n. soli of soil, the source of the organism). Cells are Gram-negative, non-spore-forming, rod-shaped (0.7–1.0 \times 1.4–2.5 μ m), containing poly- β -hydroxybutyrate or

Table 1 Differential characteristics between strain YIM 48816^T and their closest phylogenetic neighbours of the genus *Methylobacterium*.

Characteristic	1	2 ^a	3 ^a
Isolation source	Soil	Water	Leaf
Cell width (μ m)	0.7–1.1	1.0–2.2	1.1–1.6
Cell length (μ m)	1.4–2.5	2.3–4.4	1.7–5.1
Growth temperature (°C) Range	4(w)–37(w)	10–37(w)	4(w)–40
Carbon source			
L-Arabinose	—	w	+
D-Xylose	—	+	+
D-Fucose	+	w	—
D-Fructose	+	—	w
D-Glucose	+	w	w
Acetate	—	w	—
<i>N,N</i> -Dimethylformamide	w	—	—
L-Aspartate	+	—	—
Quinone composition (% of total)			
Ubiquinone Q-9	2.9	3.7	0
Ubiquinone Q-10	97.1	96.3	100.0
Fatty acid composition (% of total)			
C18 : 1 ω 7c	88.46	81.99	87.77
C18 : 1 ω 9c	0	1.10	0.57
C18 : 0	3.36	5.51	3.62
C16 : 0	3.29	8.32	3.99
C14 : 0	0.54	0.76	0.52
Sum In Feature 2	1.72	0.65	0.78
Sum In Feature 3	2.12	1.66	2.05
DNA G + C content (mol%)	66.2	69.2	69.6

Strains: 1 strain YIM 48816^T, 2 *M. mesophilicum* DSM 1708^T, 3 *M. brachiatum* DSM 19569^T.

+ Positive, – negative,

w weak

^a Data of the strains are from the parallel experiments

volutin granules. Colonies are pink to light red-coloured, circular and smooth with non-pigmented after growth on ISP 2 agar for 6 days at 28°C. Catalase- and oxidase-positive, but negative for urease activity, H₂S production, nitrate reduction, milk coagulation and peptonization, hydrolysis of gelatin, cellulose, starch and Tweens 20, 40, 60 and 80 tests. Growth is observed at 4–37°C (optimum 28°C) and pH 5.0–7.0 (optimum 7.0). No growth in the presence of NaCl concentrations of 1% or more. Enzyme activities (tested in the API ZYM system) are positive for alkaline phosphatase, esterase (C4), esterase lipase (C8), lipase (C14), leucine arylamidase, valine arylamidase, cystine arylamidase, trypsin, acid phosphatase, naphthol-AS-BI-phosphohydrolase, negative for α -chymotrypsin, α -galactosidase, β -galactosidase, β -glucuronidase, α -glucosidase, β -glucosidase, N-acetyl- β -glucosaminidase, α -mannosidase, α -fucosidase. Carbon sources utilized include D-glucose, maltose, sucrose, D-fucose, lactose, D-raffinose, D-fructose, ribose, sorbitol, myo-inositol, D-mannitol, dextrin, methanol, acetate, N,N-dimethylformamide, L-aspartate. Does not utilize L-arabinose, cellobiose, D-galactose, L-rhamnose, mannose, D-galactose, D-xylose, glycerol, calcium DL-malate. Nitrogen sources utilized include L-histidine, DL-methionine, L-leucine, L-ornithine, L-alanine, L-proline, serine, tryptophan, phenylalanine, L-valine. Does not utilize xanthine, L-arginine, urea, lysine. Ubiquinone Q-10 (97.14%) is the predominant isoprenoid quinone, the other 2.86% content is Q-9. The major fatty acid is C18:1 ω 7c (88.46%).

The type strain is YIM 48816^T (=CCTCC AA 208027^T = KCTC 22810^T), isolated from a forest soil sample in Sichuan province, Southwest of China. The DNA G + C content of the type strain is 66.2 mol%.

Acknowledgments This research was supported by the National Natural Science Foundation of China (No. 30900002 and No. 30560001), International Cooperative Key Project of Ministry of Science and Technology (2006DFA33550). Authors are grateful to Yong-xia Wang and Yun Wang for their help in the study.

References

- Austin B, Goodfellow M (1979) *Pseudomonas mesophilica*, a new species of pink bacteria isolated from leaf surfaces. Int J Syst Bacteriol 29:373–378
- Cerny G (1978) Studies on the aminopeptidase test for the distinction of gram-negative from gram-positive bacteria. Appl Microbiol Biotechnol 5:113–122
- Chun J, Lee JH, Jung Y, Kim M, Kim S, Kim BK, Lim YW (2007) EzTaxon: a web-based tool for the identification of prokaryotes based on 16S ribosomal RNA gene sequences. Int J Syst Evol Microbiol 57:2259–2261
- Cowan ST, Steel KJ (1965) Manual for the identification of medical bacteria. Cambridge University Press, London
- Felsenstein J (1981) Evolutionary trees from DNA sequences: a maximum likelihood approach. J Mol Evol 17:368–376
- Felsenstein J (1985) Confidence limits on phylogenies: approach using the bootstrap. Evolution 39:783–791
- Fitch WM (1971) Toward defining the course of evolution: minimum change for a specific tree topology. Syst Zool 20:406–416
- Gordon RE, Barnett DA, Handerhan JE, Pang C (1974) *Nocardia coeliaca*, *Nocardia autotrophica*, and the nocardia strain. Int J Syst Evol Microbiol 24:54–63
- Green PN, Bousfield IJ (1983) Emendation of *Methylobacterium* Patt, Cole, Hanson 1976 *Methylobacterium rhodinum* (Heumann 1962) comb. nov. corr., *Methylobacterium radiotolerans* (Ito, Iizuka 1971) comb. nov. corr., *Methylobacterium mesophilicum* (Austin, Goodfellow 1979) comb. nov. Int J Syst Bacteriol 33:875–877
- Kato Y, Asahara M, Goto K, Kasai H, Yokota A (2008) *Methylobacterium persicinum* sp. nov., *Methylobacterium komagatae* sp. nov., *Methylobacterium brachiatum* sp. nov., *Methylobacterium tardum* sp. nov. and *Methylobacterium gregans* sp. nov., isolated from freshwater. Int J Syst Evol Microbiol 58:1134–1141
- Kimura M (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequence. J Mol Evol 16:111–120
- Komagata K, Suzuki K (1987) Lipid and cell-wall analysis in bacterial systematics. Methods Microbiol 19:161–207
- Kumar S, Tamura K, Nei M (2004) MEGA3: integrated software for molecular evolutionary genetics analysis and sequence alignment. Brief Bioinformatics 5:150–163
- Li WJ, Xu P, Schumann P, Zhang YQ, Pukall R, Xu LH, Stackebrandt E, Jiang CL (2007) *Georgenia ruanii* sp. nov., a novel actinobacterium isolated from forest soil in Yunnan (China) and emended description of the genus *Georgenia*. Int J Syst Evol Microbiol 57:1424–1428
- Mesbah M, Premachandran U, Whitman WB (1989) Precise measurement of the G + C content of deoxyribonucleic acid by high-performance liquid chromatography. Int J Syst Bacteriol 39:159–167
- Patt TE, Cole GC, Hanson RS (1976) *Methylobacterium*, a new genus of facultatively methylotrophic bacteria. Int J Syst Evol Microbiol 26:226–229
- Saitou N, Nei M (1987) The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol Biol Evol 4:406–425
- Sasser M (1990) Identification of bacteria by gas chromatography of cellular fatty acids. USFCC Newslett 20:1–6
- Shirling EB, Gottlieb D (1966) Methods for characterization of *Streptomyces* species. Int J Syst Bacteriol 16:313–340
- Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG (1997) The CLUSTAL_X windows interface:

- flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res* 25: 4876–4882
- Xu P, Li WJ, Tang SK, Zhang YQ, Chen GZ, Chen HH, Xu LH, Jiang CL (2005) *Naxibacter alkalitolerans* gen. nov., sp. nov., a novel member of the family ‘Oxalobacteraeae’ isolated from China. *Int J Syst Evol Microbiol* 55:1149–1153